FORECASTING H. PYLORI-ASSOCIATED GASTRIC DISEASE PROGRESSION TO IMPROVE SCREENING MODALITIES FOR EARLY GASTRIC CANCER INTERVENTION

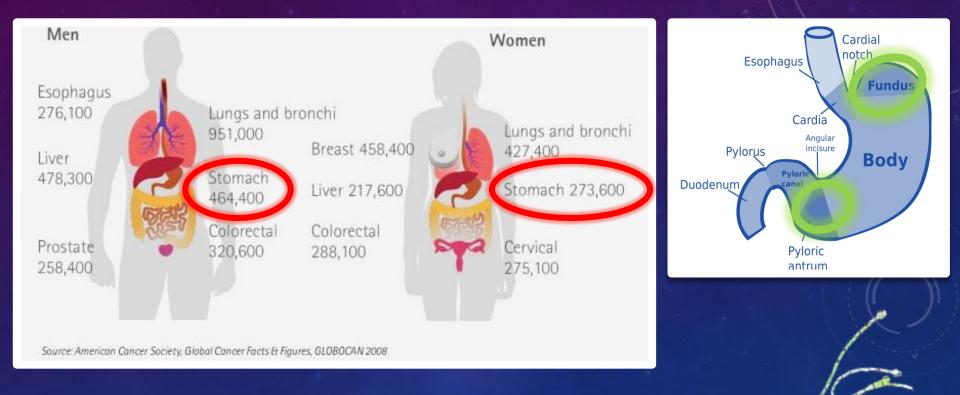
Hema, Radhia, Alicia, Derek, Ziv, Zayar, Artem, Bin, Tuhin, Yougan, Leah, Jose, Domenico, Heiko



IMO WORKSHOP 2014

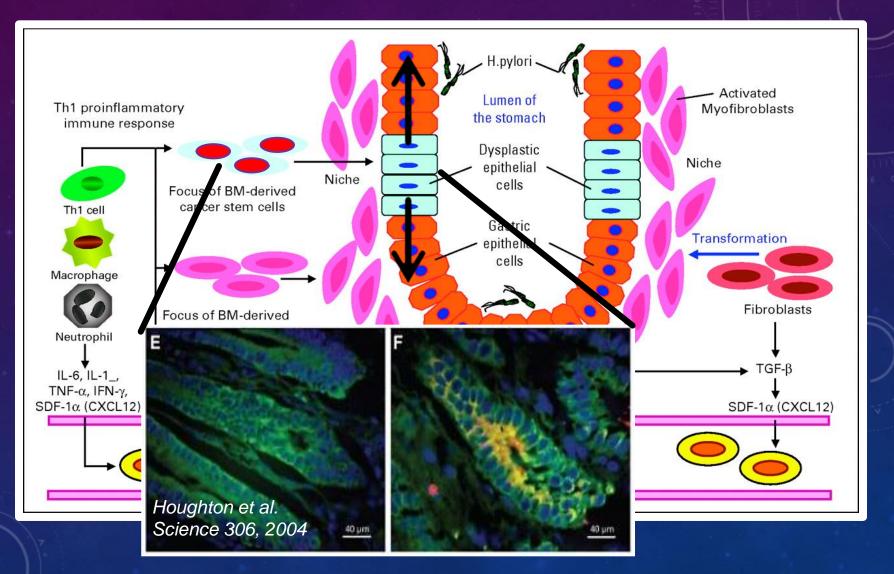


GASTRIC CANCER – 2ND MOST CANCER RELATED DEATHS WORLDWIDE



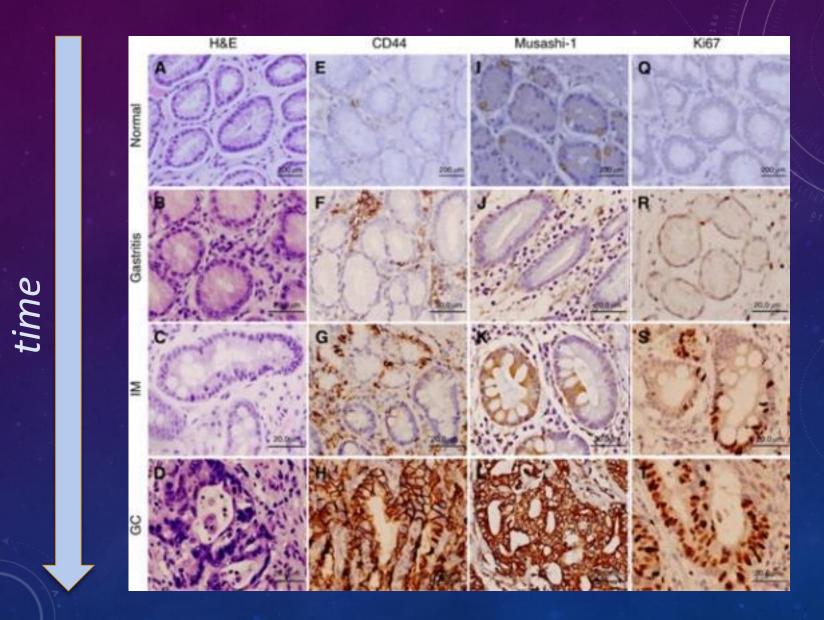
> 70% cases associated with H. Pylori

H. PYLORI IS ASSOCIATED CHRONIC INFLAMMATION THAT MOBILIZES AND RECRUITS GSCs



J Clin Oncol. 26(17), 2008

Expression of Stem Cell Markers **CD44** and **Musashi-1** increases during the progression from gastritis, IM, dysplasia and invasive cancer stages.



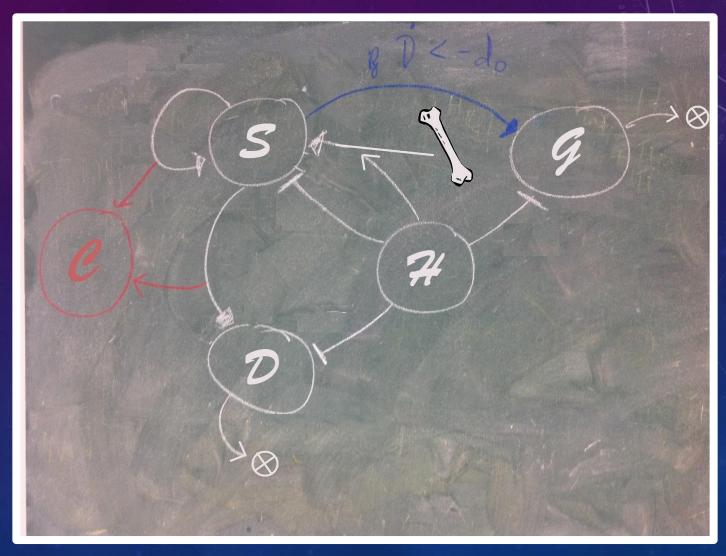
Wang et al., Br J Cancer. 105(5), 2011

<u>Hypothesis</u>

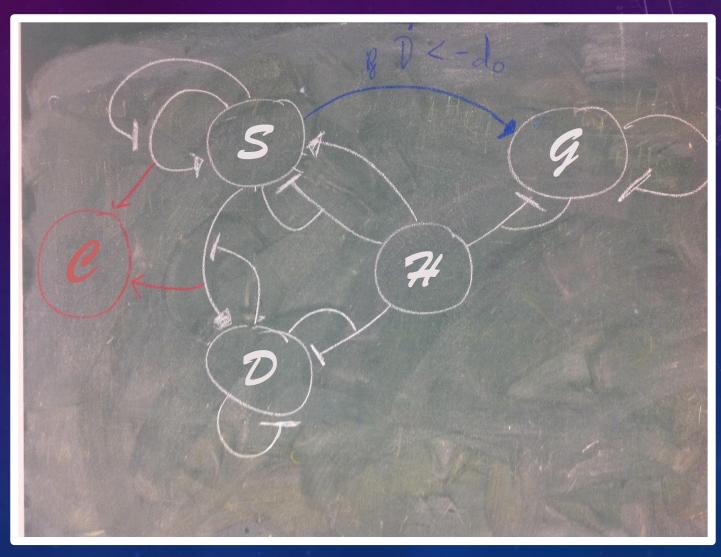
Stem cells play a pivotal role in the progression from normal to metaplasia, dysplasia, and carcinoma.

The fraction and spatial distribution of stem cells in gastric biopsies may serve as a prognostic factor for disease progression and suggest meaningful screening intervals.

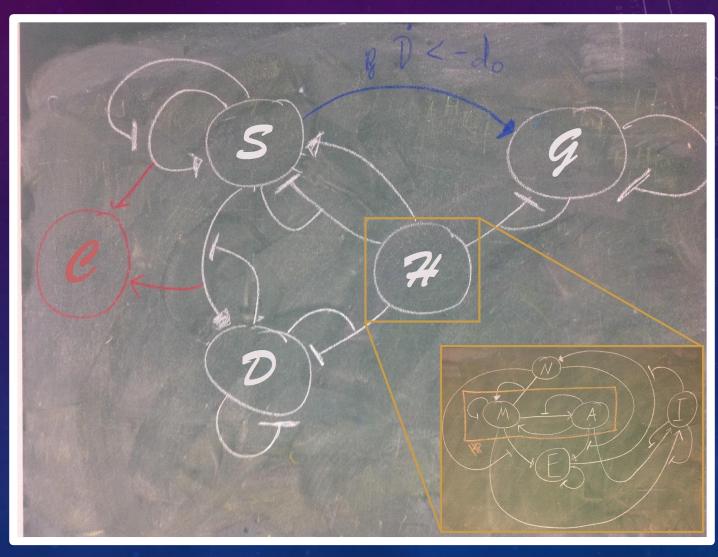
Quest: Develop a mathematical model of H. Pylori-associated Carcinogenesis



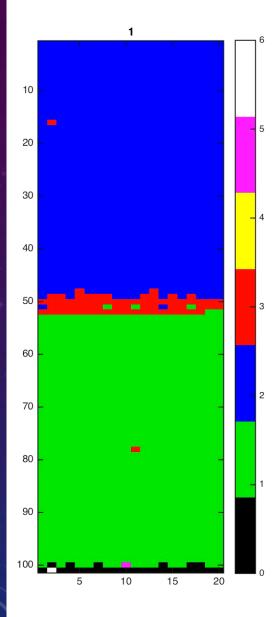
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Agent Based Model



Dysplasia/Cancer

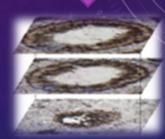
Goblet Cells

Gastric Stem Cells

Mucosal Cells

Secretory Cells (Chief, Parietal, G Cells)

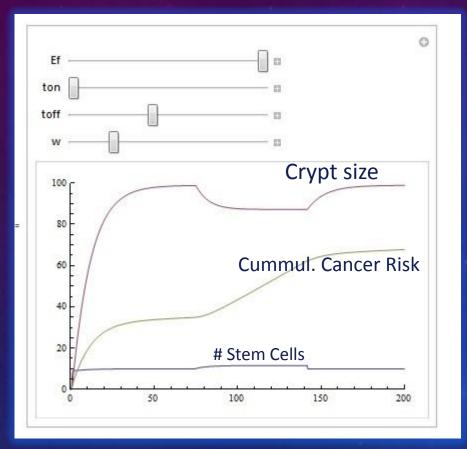


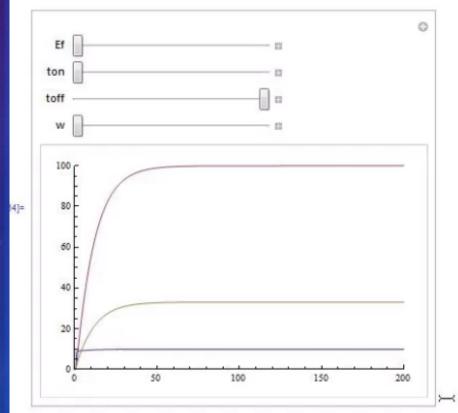


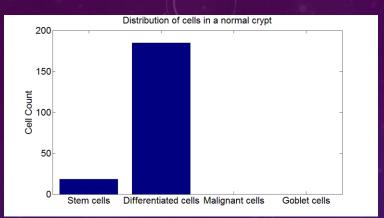


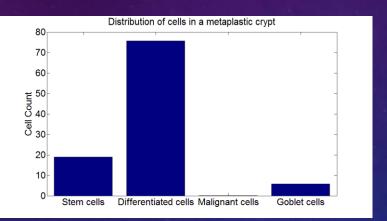


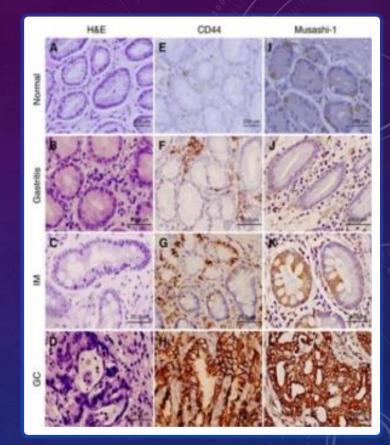
Crypt Dynamics

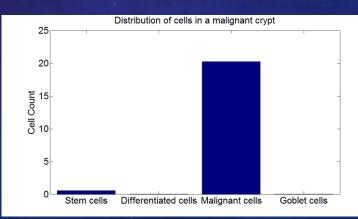












AIM 1.

Develop a *mechanistic mathematical* model of gastric crypt homeostasis and *H. Pylori* induced inflammation and facilitation of carcinogenesis.

Calibrate the model with <u>stem cell</u> numbers and their spatial distribution at <u>different gastric disease stages</u> with retrospective tissue data from <u>H. Pylori-associated disease</u> (Cali, Columbia; n=30) and <u>non H. Pylori-associated</u> disease (Moffitt; n=30) provided by Dr. D. Coppola.

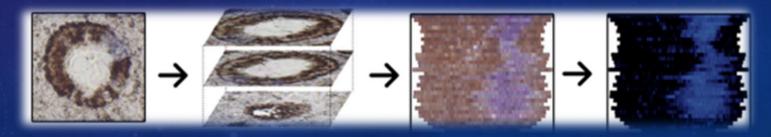


Gastric biopsy tissue samples from different disease stages (normal, hyperplasia, metaplasia, cancer)

- H. Pylori-associated; Cali (n=30)
- Non H. Pylori-associated; Moffitt (n=30)

To be done: staining for GSC: CD44, Musashi-1

Quantification by **Aperio system** in the Analytic Microscopy Core at MCC to count the positive GSCs and to correlate their number to the surface area evaluated per tissue sample.



AIM 2!

Use the calibrated model to *predict* patient-specific disease progression dynamics using <u>sequential screening samples</u> from endoscopic gastric biopsies (n=10; Dr. D. Coppola).

We will randomize the retrospective data into <u>training and test cohorts</u> to validate the predictability of the disease progression model to suggest personalized screening schedules for early intervention.

AVAILABLE DATA

Single patient (n=10) gastric biopsy tissue samples from different disease stages (normal, hyperplasia, metaplasia, cancer)

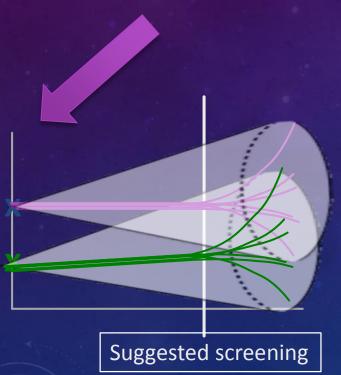
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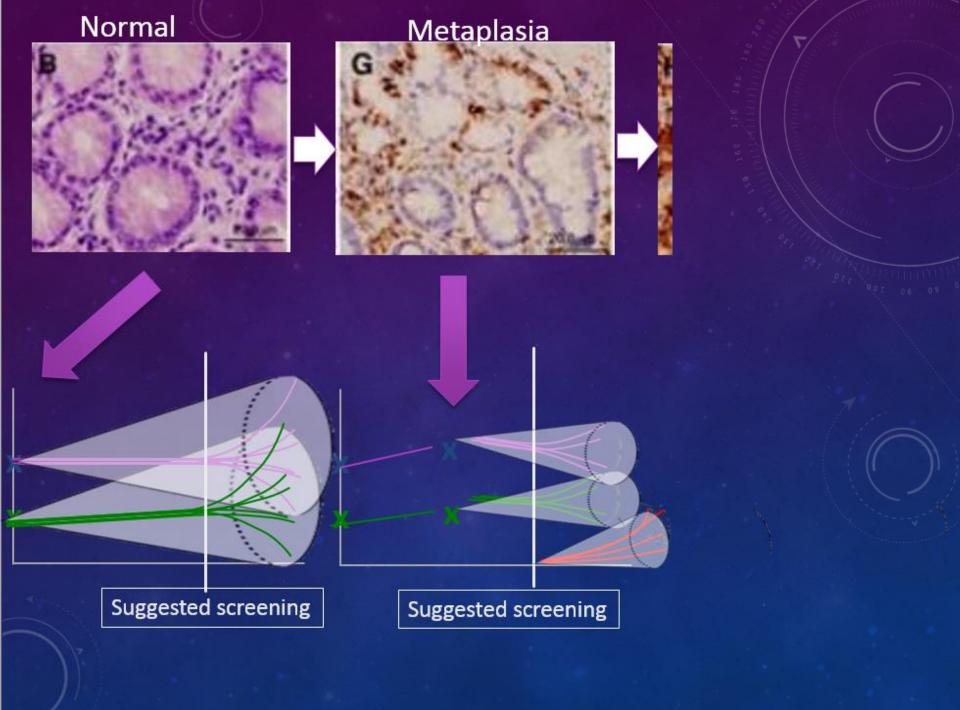
MODEL DESIGN

- Collect patient specific data
- Use as initial condition for math model
- Use derived <u>parameter distributions</u> to predict disease progression within 'cone of uncertainty'
- Use subsequent patient samples to re-calibrate the model and forecast disease progression with a smaller 'cone of uncertainty'

Normal







Normal

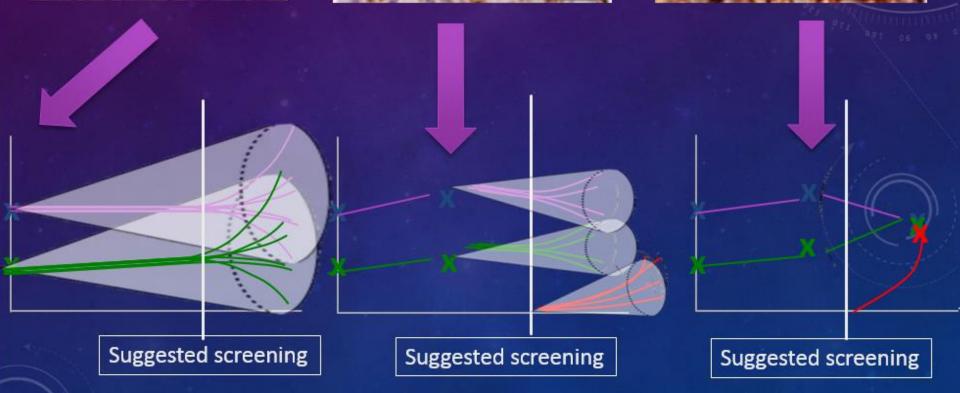


<u>Metaplasia</u>

G

Low Grade Dysplasia



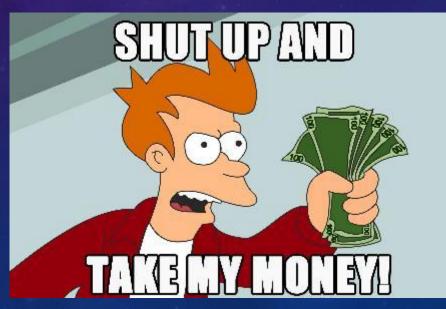


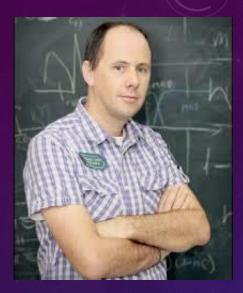
SUMMARY

- We are proposing an integrated mathematical model parameterized by clinical samples to identify H. pylori infected patients who are at greater risk to develop gastric dysplasia and carcinoma
- The model, initialized to patient-specific data, will suggest personalized screening schedules

<u>BUDGET</u>

- IHC on available retrospective data \$20,000
- 50% effort Postdoc for model calibration and validation \$30,000







THANK YOU.





